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Observational Evidence for Teleportation of Living Systems between Sealed Compartments

[1]Benjamin J. Scherlag, PhD, [1]Khaled Elkholey, M.D,
Jerry I Jacobson, PhD, DMD

[1]The Department of Medicine and The Heart Rhythm Institute, The University of Oklahoma Health Sciences Center, Oklahoma City, OK

[2]Institute of Theoretical Physics, Jupiter, FL

Corresponding Author: Benjamin J. Scherlag, PhD)
800 Stanton L Young Blvd, Suite 5400, Oklahoma City, OK 73104, USA.

Abstract

This study describes the teleportation of living organisms. Methods: Culture media containing the unicellular animal, Stentor coeruleus, placed in uncovered deep well slides (n=22), was allowed to evaporate for 24 hours (Group 1). After adding tap water (pH=7.5) and sealing the deep well slides with a snap cover, cells showing disrupted membranes released their micronuclei. Each slide was covered by another sealed deep well slide containing sterile Stentor media (pH =6.9, Group 2). A plastic collar (width, 8 mm) was placed on the first deep well slide and the added second slide formed an air space between slides (Group 3, n=10). Deep well slides were filled with previously boiled Stentor media and sealed (Group 4, n=5). All slides were viewed microscopically daily for 10 days. Results: In the deep well slides of Group 1, only 1 of 22 showed micronuclei which acted as progenitors splitting into 4 mobile dwarf cells; whereas, in Group 2, 7/22 experiments, showed both progenitor and dwarf cells, $p < 0.05$, compared to Group 1. In the 10 experiments with an air space between the two sealed slides (Group 3), 20 % showed progenitor and mobile dwarf cells. The sealed deep well slides containing the sterile Stentor media (Group 4) showed neither progenitor nor dwarf Stentors. Conclusions: We found that progenitor cells and dwarf forms which developed from the micronuclei of the larger varieties of Stentor coeruleus cells migrated from one sealed compartment containing an unfavorable environment to another sealed compartment containing a more favorable environment.

Keywords: Stentor coeruleus, Teleportation, Progenitor cell, Dwarf Stentors, Quantum Tunneling.

INTRODUCTION:

Stentor microorganisms, sometimes called trumpet animalcules, are a group of unicellular, filter-feeding, ciliate protozoans. They are usually horn-shaped, but can take other forms, i.e., ovoid, and reach lengths of a millimeter. Stentor coeruleus has commonly been used in experimental settings because they can, after being surgically cut into fragments, regenerate into a fully formed organism [1]. Some Stentor species react to unfavorable environmental conditions by contracting into an encysted or vegetative state.

In the present study we noted that, when the liquid medium was allowed to evaporate, some encysted Stentor cells manifested disrupted membranes allowing the release of multiple micro-nuclei [2, 3]. These micronuclei, when examined microscopically, showed rapid movement of their internal inclusions and could act as progenitor cells dividing into 4 mobile dwarf forms [4]. In the present study, we found that these progenitor cells and dwarf forms could migrate from one sealed compartment containing an unfavorable environment to another sealed compartment containing a more favorable environment over a period of 24-48 hours.

From a theoretical standpoint, it is proposed that the micronuclei which appeared to have electromagnetic properties can act in accordance with the Jacobson Resonance Theory, based on the equation $Mc^2=BvLq$, where m is the mass of a particle in the 'box' or 'string' (molecule in a biosystem), c is the velocity of electromagnetic field in space, independent of its inertial frame of reference, B is the magnetic flux density, v is the velocity of the carrier or 'string' (a one or two dimensional 'box') in which the particle exists, L is its dimension (length) and q represents a unit charge q=1C. By defining electromotive force as energy per unit charge, equivalencies suggest that qvBL is one of the fundamental expressions of energy of a charged wave-particle in magnetic fields. The micronuclei disappeared as virtual particles, whereas some could reappear ostensibly through quantum tunnelling, as progenitors produce dwarf stentors in the favorable environment; overcoming the frank quantum barrier. We believe that this demonstration of teleportation of a unicellular organism, wherein the functional nature of life was maintained; may also occur at the multicellular level.

METHODS

Uncovered deep well slides (Fisher Scientific. Waltham, MA, Figure 1) were filled with culture media containing the protozoan, Stentor coeruleus (Carolina Biological Supply, Burlington, NC). On standing for 24 hours the liquid dried and many encysted microorganisms were viewed microscopically (Group 1, n=22) Many of the large ovoid cells showed disrupted membranes. Each deep well slide was then filled with tap water (pH=7.5) and the cover snapped shut to seal the contents.

A second set of deep well slides (Group 2, n=22) were filled with previously boiled and filtered Stentor media (pH=6.9) and sealed with a snap cover. The deep well slides containing the sterile media were placed on the first to form a set (Figure 2).

Each sealed deep well slide of Groups 1 and 2 was examined microscopically daily to determine the presence of active cells, i.e., progenitor or mobile dwarfs. Still microphotographs and videos were recorded at magnification between 10X and 60X.

In ancillary experiments, a plastic collar (width, 8 mm) was placed on the first deep well slide and the second slide placed so that there was no direct contact since there was an air space between slides. (Group 3, n=10). A control set of deep well slides were filled with previously boiled Stentor media and sealed (Group 4, n=5). These slides were removed to another room and viewed microscopically 24-48 hours later as well as up to 10 days later.

Definition of Terms:

Teleportation. The transfer of matter or energy from one point to another without evidence of movement across the physical space between them.

Dwarf Stentor. Mobile unicellular organisms, 25-40 microns in diameter that derived from micronuclei released from the large variety of Stentor coeruleus (400-1000 microns in diameter) whose membranes were damaged during evaporation of the culture media.

Statistical Analysis

Analysis was performed using a Chi square test (2 x 2 contingency table) followed by a 2 tailed Fisher exact test comparing the number of cells showing progenitor cells displaying internal movement of their inclusions and the presence of mobile dwarf Stentors between Groups 1 and 2. A p value of < 0.05 was considered significant.

RESULTS

Figure 1 shows an opened deep well slide to the right and the snap cover on the left. The diameter of the well measures 3 cm.

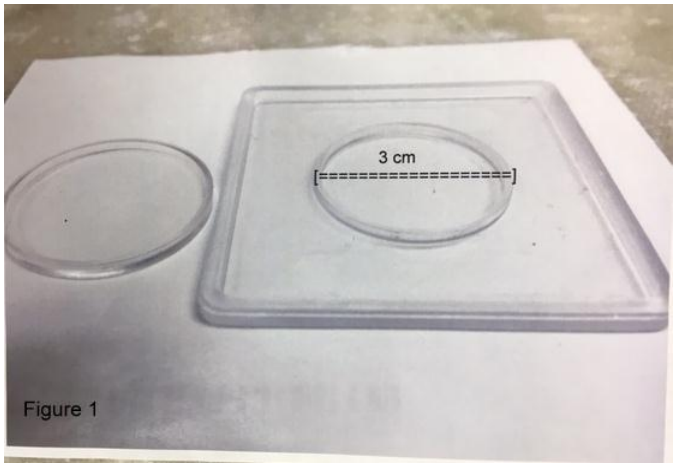

Figure 1. An illustration of an opened deep well slide (on the right) and the snap cover (at the left). The diameter of the well is 3 cm, capacity: 0.14 cc.

Figure 2 shows sealed deep wells slides, placed on each other. Group 1 contained the hydrated Stentor cells and the sealed deep well slide containing the sterile Stentor media placed on top of each other, Group 2.

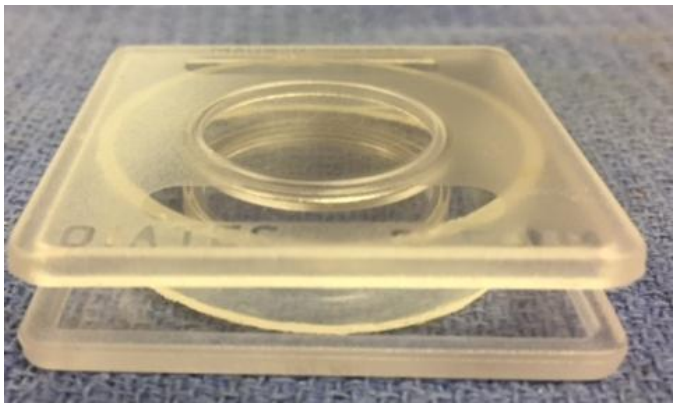

Figure 2. The positioning of one sealed deep well slide, containing sterile Stentor media placed onto a sealed deep well slide containing the evaporated contents of the Stentor encysted cells hydrated with tap water.

Figure 3. There are a variety of the large cell forms of Stentor. Another form is oblong in shape. At the buccal end 2 sets of cilia propel food (small microorganisms) into the body cavity. All forms have a set of active cilia surrounding the "mouth".

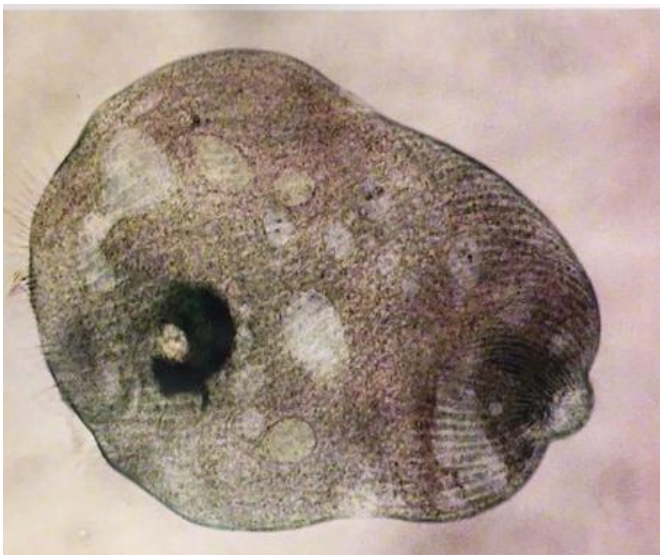

Figure 3. A typical illustration of an ovoid Stentor coeruleus organism. Note the cilia at the buccal end and the characteristic stripes. Cell sizes range from 750-1000 µm

Figure 4 depicts an example of an ovoid encysted cell, 24 hours after the culture media had evaporated. Note that a section of the membrane has been disrupted allowing release of micro-nuclei (arrows) into the local environment. The deep well slides were then hydrated with tap water and the slide sealed with its plastic cover.

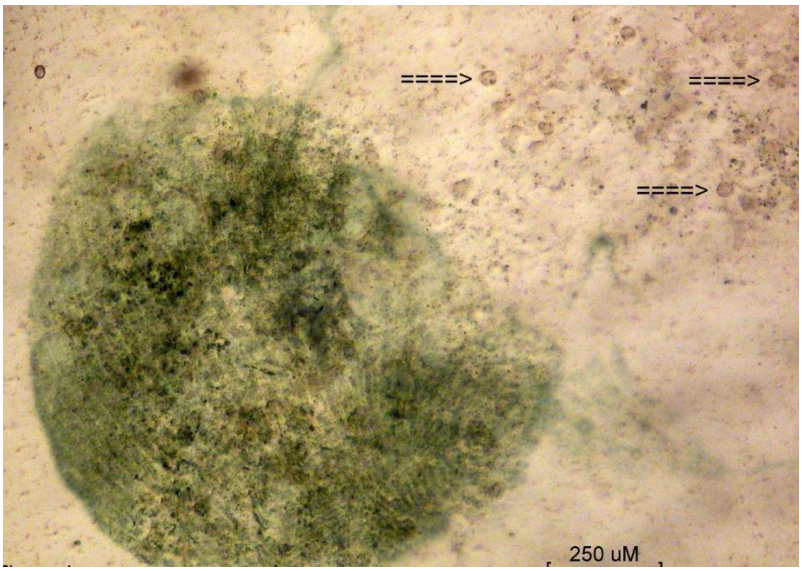

Figure 4. Disrupted void Stentor cell whose membrane rupture on dehydration releasing micronuclei (arrows). Bar 250 microns.

Figure 5, video, CTRL/left-click on link below or copy/paste into browser (https://www.dropbox.com/s/fo7tmo4jm8ebmav/PICT0077.AVI?dl=0) shows three micro-nuclei that migrated to the sealed compartment containing the sterile Stentor media. Note that all these cells manifest different degrees of actively rotating inclusions. The cell with the most activity starts to segment and splits off into 4 dwarf cells which separate and swim away. (move to time to 2:40, min:secs, respectively)

Click the below link to see the Figure 5 (a video)

https://www.dropbox.com/s/fo7tmo4jm8ebmav/PICT0077.AVI?dl=0

Figure 5, video. Three micronuclei turned into active progenitor cells with rotation of their internal inclusions. The cell on the left shows segmentation and splits off into 4 dwarf cells which separate and swim away.

In the deep well slides of Group 1, careful examination of these preparations revealed only 1 of 22 in which we found an active progenitor cell. In contrast the deep well slides

which were filled with the sterile Stentor Media (Group 2) 7 of the 22 (32%) showed active progenitor cells, many of which were actively dividing into mobile dwarf Stentors. Indeed the deep well slides in Group 2 were teeming with mobile dwarf Stentors. Thus, the presence of active progenitor cells and mobile dwarf cells in the sealed compartments of Group 2 were significantly greater than in the sealed deep well slides of Group 1 (<0.05).

Controls:

The sealed deep well slides containing the sterile Stentor media that were set aside as controls showed neither progenitor cells nor dwarf Stentors in the same 24 hour period or during the subsequent 10 days of examination.

Ancillary experiments:

In the 10 experiments in which a plastic collar was interposed between the two sealed deep well slides of Group 3 and Group 4, 20 % of the latter showed active progenitor cells and mobile dwarf Stentors compared to none of Group 3.

DISCUSSION

Major Findings:

In the present study, under microscopic examination, the damaged membranes of the encysted forms of Stentor coeruleus were shown to release micro-nuclei which displayed movement of their internal inclusions. These micro-nuclei migrated from one sealed compartment to another before acting as progenitors by dividing into two or, more commonly, four mobile dwarf Stentor organisms.

Background:

There have been 2 major treatises exclusively devoted to the Genus Stentor, particularly to the species coeruleus which were authored by Johnson [2] and Tartar [3]. Both mention the appearance of diminished forms of the larger varieties due to starvation. Under these circumstances, the organisms self-cannibalize their internal contents. The starved forms are reduced in size ranging from 94-376 microns but regain their previous size when fed. These descriptions contrast with the dwarfs seen in the present study. These dwarfs ranged in size from 30-45 microns and derived from dividing micronuclei shed from the disrupted encysted cell of the much larger forms (Figure 4). Moreover, although there was a small disparity in the relative sizes of these dwarfs they did not grow over several days of observation. As described previously, after several days, assuming the diminution of nutrients, we observed the encysting of these dwarf cells which characteristically formed into clusters with loss of their internal inclusions (Figure 5)

In regard to the micronuclei within the Stentor cells, Johnston's described them as "spherical, highly refractive, deep-staining bodies measuring 1.5-2 microns in diameter apparently homogeneous and composed wholly of chromatin." These greatly enlarge at the time the organisms show reproduction by fission or conjugation. These observations have been confirmed by a later study [5] which found that the micronuclei of Stentor

coeruleus enlarged to >20 microns during mitosis. However, there is no mention of four dwarfs arising from a single micronucleus as we found in micronuclei released from the disrupted Stentor cells. Those measured 20-30 microns in diameter. It is important to note the consistent type of activity observed in the present study before and during division of the micronuclei/progenitor cells. Specifically, the inclusions within the cell interior progressively increased their rotation before cell segmentation into 2, or more commonly, 4 dwarfs which separated before swimming away. This scenario of an unusual and striking form of cell division that could not be found in careful reviews of the monographs of either Johnson [2], Tartar [3] or Raikhel et al. [5].

Teleportation Mechanisms

Possible explanations could be proposed to explain the apparent migration of the micronuclei/progenitor cells across the plastic barriers (Figure 5, video) and the interposed air space described in the present report:

1. The cells could deform themselves so as to move through the pores of the plastic covers of the deep well slides. A survey of the internet found that the pores of the polystyrene material comprising the deep well slides are 0.4 microns in diameter. It seems unlikely that these cells could be able to navigate through such pores 2 orders of magnitude smaller than their body dimensions through one layer of plastic let alone two. In the experiments with the interposed air space this explanation becomes even less plausible.
2. Another alternative is that the bioelectromagnetic nature of the Stentor micronuclei allows them to migrate from an unfavorable environmental (tap water, pH=7.5) to a more favorable media (sterile Stentor media, pH=6.9). In a previous report, we demonstrated that replicate images of plant leaf structures could be retrieved through glass using magnetized nano-sized iron particles and a Prussian Blue iron staining solution [6, Figure 3]. More recently, we demonstrated that, using a similar imaging technique, replicates of animal tissues could be imaged through glass or other physical barriers [6-8].

IMPLICATIONS

The concept of teleportation, as a physical phenomenon, has only recently been shown in experiments in which subatomic particles could be teleported across space in accordance with quantum theory [9, 10]. In quantum law, particles can exist simultaneously in two different states. These principles are known as superposition and entanglement and provide the basis for sub-atomic particle teleportation. We hypothesize that living organisms exist in two different superposition and entangled states, physical and electromagnetic. Under specific circumstances, e.g., unfavorable and favorable environmental conditions, these biological states can separate and the electromagnetic state can be subject to teleportation.

LIMITATIONS

These experiments were not conducted using solutions with different types of media with different pH levels to determine potential enhancing or inhibitory effects on progenitor or dwarf cell formation. Tartar in 1957 [6] applied different solutions and chemicals to cultures of Stentor coeruleus to determine effects on"cytoplasmic differentiation" and morphogenesis. It is interesting to note that Tartar found toxic effects on Stentor when he exposed them to sea water. It should be pointed out that a recent review [11] of Stentor coeruleus in areas of study ranging from regeneration to associative behavior [12] make this organism a useful model for ongoing experimental attention as evidenced by the present study.

CONCLUSIONS

We found that progenitor cells and dwarf forms which developed from the micronuclei of the larger varieties of Stentor coeruleus cells could migrate from one sealed compartment containing an unfavorable environment to another sealed compartment containing a favorable environment.

Furthermore, it is conceivable that quantum tunneling occurred based upon possible accommodative alterations in the electromagnetic field environment. Relative translational motion of biological matter, ultimately comprised of elementary electric charges, could move into a different state of reality. It is theorized further, that quantum tunneling may be achieved via electromagnetic field regulation of baryons and mesons to therein provide gluon transmissions, i.e. the strong nuclear force, wherein quark domains were regulable via Higgs bosonic interactions.

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